SYNTHESIS OF GLUCOSYL PHOSPHATIDYLGLYCEROL VIA A PHOSPHOTRIESTER INTERMEDIATE.

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Complete deblocking of a fully-protected intermediate, which was prepared by using *inter* alia a selective protection of 1- or 3-0-ally1-sn-glycerol with one pivaloyl group and effective phosphorylation and coupling procedures, afforded glucosylphosphatidylglycerol.

It is well known now<sup>1)</sup> that glucosylphosphatidylglycerol (i.e., <u>8d</u>) is part of the phospholipid pool of moderately halophilic halotolerant Gram-negative bacteria.

In order to get more insight into the biosynthesis and function of naturally occurring glycophospholipids we embarked upon a programme to develop efficient synthetic methods for the preparation of this type of compounds.

In this paper we wish to present a convenient and attractive procedure for the synthesis of glucosylphosphatidylglycerol (8d).

The compound to be synthesized contains two ester bonds, one  $\alpha$ -glucosidic linkage and one phosphodiester linkage. In order to introduce the latter two types of linkages, and keeping in mind the base lability of the two ester bonds, we adopted the following strategy. The  $\alpha$ -linkage between D-glucose and sn-glycerol was introduced by condensing 2,3,4,6tetra-0-benzyl-D-glucosyl bromide<sup>2)</sup> (1,  $R^{l}$ =benzyl) with the secondary hydroxyl group of a suitably protected sn-glycerol derivative (i.e., 2) under the conditions as developed by Lemieux<sup>3)</sup>. For the protection of the two primary hydroxyl groups of sn-glycerol we selected the allyl and the pivaloyl groups. The choise of the latter relatively base stable 4) group had several attractive features. For instance, under the conditions of Lemieux, which are necessary for the required a-glucosidic linkage, no migration of this group was observed, further, the removal of this group could be accomplished selectively to give a free primary hydroxyl function which was necessary for the formation of the required phosphoester bond with the phosphatidic derivative (7). Another noteworthy feature adherent to the use of the pivaloyl group was that it could be introduced selectively at a primary hydroxyl of a snglycerol derivative. Thus treatment of 1-0-ally1-sn-glycerol<sup>5)</sup> (15 mmol) with pivaloy1 chloride (15 mmol) in dry pyridine at  $-10^{\circ}$ C gave, after purification by short column chromatography<sup>6)</sup>, pure  $2^{7a,b}$  (R<sup>2</sup>= (CH<sub>3</sub>)<sub>3</sub>CCO ; 13.8 mmol). In the same way, starting from 3-0ally1-sn-glycero1<sup>5)</sup>, the enantiomeric form of 2 was obtained. The identity of both compounds was established by  $^{1}$ H-NMR and  $^{13}$ C-NMR spectroscopy.

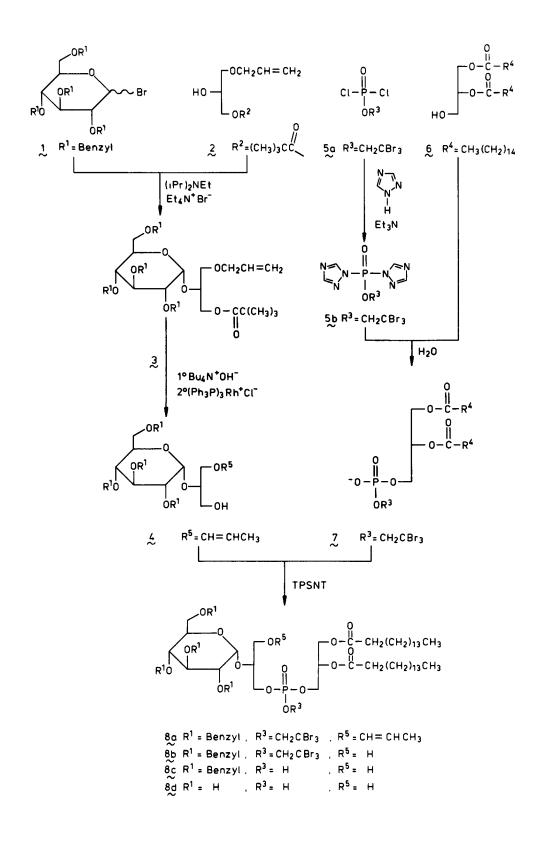
Condensation of the glucopyranosyl bromide <u>1</u> (R<sup>1</sup>=benzyl ; 3.5 mmol) with the sn-glycerol derivative <u>2</u> (2.5 mmol), under the conditions of Lemieux, afforded, after purification by silica gel chromatography, the required product <u>3</u> (1.2 mmol) as a homogeneous oil ( $(\alpha)_D^{25}$ = +31.6°, c=1, CHCl<sub>3</sub>). In the same way, starting from 1-0-pivaloy1-3-0-ally1-sn-glycerol, an other diastereomer of <u>3</u> ( $(\alpha)_D^{25}$  = +38.8°, c=1, CHCl<sub>3</sub>) was obtained. The two compounds thus obtained could also be distinguished by <sup>1</sup>H-NMR spectroscopy. Thus the anomeric proton H1 of the glucose part of the 1-O-allyl derivative of 3 had a  $\delta$ -value of 5.05 (d, J = 3.1 Hz) and the 3-O-allyl derivative of 5.18 (d, J = 3.1 Hz).

Treatment of a solution of 3 (0.73 mmol) in dioxane-water with tetrabutylammonium hydroxide for 4 h at 20°C gave, after work-up and purification by short column chromatography,  $4^{7a,b}$  (R<sup>5</sup>= CH<sub>2</sub>CH=CH<sub>2</sub>; 0.64 mmol) as a homogeneous syrup.

The derivative  $\underline{4}$  (R<sup>5</sup>= CH=CHCH<sub>3</sub>) necessary for the coupling with the phosphatidic derivative  $\underline{7}$  was obtained by treating  $\underline{4}$  (R<sup>5</sup>= CH<sub>2</sub>CH=CH<sub>2</sub>; 0.5 mmol), in ethanol/water (9:1,v/v) containing 1,4-diazabicyclo(2,2,2) octane (20 mg), with (Ph<sub>3</sub>P)<sub>3</sub>RhCl<sup>8</sup>) (45 mg) for 2.5 h at 85°C. Work-up of the reaction mixture followed by short column chromatography afforded  $\underline{4}$  (R<sup>5</sup>= CH=CHCH<sub>3</sub>; 0.48 mmol) as an oil. Analysis of this oil by <sup>1</sup>H-NMR spectroscopy revealed the presence of  $\underline{4}$  (R<sup>5</sup>= CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>; <u>ca</u>. 10%). The latter impurity which accompanies<sup>9</sup>) the isomerization of the allyl group into the prop-1-enyl group could easily be removed in the last step of the synthesis of the crucial intermediate <u>8b</u>.

The last steps involved in the synthesis of 8d consisted of the introduction of a phosphotriester linkage between the derivative 4 and the phosphatidic derivative 7 followed by a stepwise and selective removal of all protecting groups. The synthesis of 1,2dipalmitoyl-sn-glycero-3-phospho-2,2,2-tribromoethanol (7) was accomplished by phosphorylation of 1,2-dipalmitoyl-sn-glycerol<sup>10)</sup> (6) with the in situ prepared 2,2,2-tribromoethylphosphoroditriazolide (5b). The latter phosphorylating agent was prepared 11) by reacting together equimolar amounts of 5a<sup>12)</sup> in THF with triazole in the presence of triethylamine. A solution of 6 (0.35 mmol) in dry pyridine was added dropwise to crude 5b (0.53 mmol) which was obtained after filtering off triethylamine hydrochloride. After two hours, when TLC analysis (CHCl<sub>2</sub>/MeOH/H<sub>2</sub>O, 80:20:1,v/v) showed the reaction to be complete. The reaction mixture was worked-up and product 7 was isolated by silica gel chromatography to afford, after conversion of compound 7 into the triethylammonium salt by extraction of a solution of the compound in chloroform with triethylammonium bicarbonate (2 M, pH 7.5), 7<sup>7b)</sup> (0.31 mmol) as a waxy compound. It is noteworthy to mention that the above described phosphorylation procedure, despite the use of the bifunctional phosphorylating agent 5a, gives no rise to the formation of symmetrical products.

Coupling of  $\frac{4}{2}$  (0.4 mmol) with  $\frac{7}{2}$  (0.46 mmol) to give  $\underline{8a}$  was effected in dry pyridine under the influence of the coupling agent 2,4,6-tri-isopropylbenzenesulphonyl-3-nitro-1,2,4triazolide<sup>13)</sup> (TPSNT; 0.48 mmol). After 2 h at 20°C, the crude reaction mixture was workedup and purified by short column chromatography, to give compound  $\underline{8a}$  (0.33 mmol) which was contaminated with <u>ca</u>. 10% of <u>8a</u> (R<sup>5</sup>= CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>). Treatment of <u>8a</u> (0.25 mmol) with HgCl<sub>2</sub>/ HgO<sup>14)</sup> in acetone during 20 min. at 20°C gave, after work-up and purification by short column chromatography, <u>8b</u> (0.19 mmol) as a viscous oil ( $\alpha_{D}^{25}$  = +19.7°, c=1, CHCl<sub>3</sub>) which was now devoid of the impurity introduced during the isomerization of the allyl group into the prop-1-enyl group (i.e., isomerization of <u>3</u> into <u>4</u>). Removal of the 2,2,2-tribromoethyl group from the phosphotriester <u>8b</u> to give <u>8c</u> was easily accomplished by treating a solution of <u>8b</u> in pyridine with zinc dust which was activated by the addition of a few drops of pentane-2,4-dione<sup>15)</sup>. After stirring the reaction mixture for 10 min at 40°C, followed by work-up of the reaction mixture, <u>8c</u> was obtained as a light yellow oil. ( $[\alpha]_{D}^{25}$  = +29.7°,



c=1, CHCl<sub>3</sub>). The homogeneity and identity of  $\underline{8c}$  was established by TLC-analysis, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR and <sup>31</sup>P-NMR spectroscopy.

Conversion of <u>8c</u> into <u>8d</u> was accomplished by hydrogenation of <u>8c</u> (0.095 mmol) in isopropanol/ethylacetate/acetic acid (6:3:1,v/v) over palladium on charcoal for 2 days at  $20^{\circ}$ C. Work-up of the reaction mixture and fractionation by short column chromatography, followed by conversion of the compound into the triethylammonium salt, gave pure <u>8d</u> (0.065 mmol) as a waxy solid. The identity and homogeneity of <u>8d</u> was unambiguously confirmed by <sup>1</sup>H-NMR, <sup>13</sup>C-NMR and <sup>31</sup>P-NMR spectroscopy.

In conclusion, the methodology described in this paper presents an elegant way for the synthesis of naturally occurring glycophospholipids. For instance, derivative  $\underline{8b}$  is a very convenient intermediate for the synthesis of a recently<sup>16)</sup> isolated glucosylated diphosphatidylglycerol from B Streptococci.

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## FOOTNOTES AND REFERENCES

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